

• 综述 •

库普弗细胞在非酒精性脂肪性肝病发病机制中的作用

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摘要:库普弗细胞(KC)是门脉系统的第一道屏障,也是机体固有免疫系统的重要组成部分,能分泌大量的炎性介质等物质,参与多种肝脏疾病的发生发展。非酒精性脂肪性肝病是指除乙醇等明确的损伤因素外,以弥漫性肝细胞大泡性脂肪变为病理特征的临床综合征,是仅次于病毒性肝炎的常见肝病。近年来研究发现 KC 与该病的发病全过程密切相关。

关键词:库普弗细胞;非酒精性肝病;机制

库普弗细胞(KC)是肝脏内固有巨噬细胞,长期以来人们一致认为它是清除门脉系统特定物质如内毒素等的主要细胞。然而,近年来越来越多的证据提示 KC 还参与了多种肝脏疾病的发生、发展,如:病毒性肝炎、酒精性肝病、肝内胆石症、脂肪肝、肝纤维化、以及激活或抑制肝移植中的排斥反应等。本文对近年来 KC 在非酒精性脂肪性肝病(NAFLD)发病机制中的作用研究进展作一综述。

1 KC 概述

作为机体固有免疫系统的重要组成部分, KC 可以是直接发挥效应的细胞,也可以作为抗原提呈细胞或多种细胞因子的重要来源,发挥它在机体固有免疫、抗感染等多方面的作用。同时, KC 是胃肠道吸收物质入血后面临的第一道屏障,能清除微生物、内毒素、免疫复合物、老化细胞和毒素(如乙醇等),同时由于 KC 分布位置的重要性,它还可作为抗原提呈细胞参与肝脏的再生与肿瘤发生。KC 还能分泌大量的可溶性炎性介质,在机体固有免疫和机体防御功能中起关键作用。现有大量证据表明: KC 与脂多糖(LPS)的相互作用是多种疾病中导致肝细胞损害的始动因素^[1],在多种肝脏损伤如:内毒素血症、缺血再灌注损伤、病毒感染、酒精性肝病和 NAFLD 的过程中都起到了重要的作用。

2 KC 与 NAFLD

NAFLD 是指除乙醇和其他明确的肝损伤因素外,以弥漫性肝细胞大泡性脂肪变为病理特征的临床综合征,包括单纯性脂肪肝(SFL)以及由其演变的脂肪性肝炎(NASH)和肝硬化(FLC)三种类型,是仅次于病毒性肝炎的常见肝病。大多数 NAFLD

病例发生于肥胖、高脂血症、糖尿病患者,胰岛素抵抗、氧应激、细胞因子可能是其发病环节中的核心因素。近来研究还发现 KC 与非酒精性肝脏脂肪变性、肝炎、肝硬化的发病全过程密切相关。

2.1 KC 与肝细胞脂肪变性

2.1.1 NAFLD 与胰岛素抵抗 现有研究表明,几乎所有的 NAFLD 患者都存在肝脏和脂肪组织的胰岛素抵抗(IR), IR 的严重程度与 NAFLD 的病情进展呈正相关。IR 使血清中游离脂肪酸(FFA)增多,当过量的 FFA 超出肝脏的代谢能力时,就会导致肝细胞脂肪变性。目前已基本公认 NAFLD 是代谢综合征(IR 综合征)的表现之一^[2-4]。

2.1.2 KC 与 NAFLD 胰岛素抵抗 研究发现炎性反应可增加 IR^[2]。作为肠道吸收物质入血后的第一道屏障, KC 可被 LPS、饱和脂肪酸、高糖等多种物质激活,引发肝脏局部甚至全身的炎性反应,参与 IR。

实验表明, KC 是机体特别是肝脏局部 TNF- α 的主要来源^[5], 而 TNF- α 是调节胰岛素敏感性的重要细胞因子。研究发现,在 NASH 患者的脂肪组织和肝组织内 TNF- α mRNA 表达明显上调。一方面, TNF- α 能直接抑制调节脂肪酸代谢的蛋白表达如脂肪酸结合蛋白 aP5、葡萄糖转运子 4(GLUT4), 从而影响胰岛素发挥正常生物学功能, 或者直接磷酸化胰岛素受体底物 1(IRS-1)的丝氨酸位点, 并将 IRS-1 转化为胰岛素受体酪氨酸激酶的抑制剂, 从而减少胰岛素受体的酪氨酸激酶活性^[6], 影响胰岛素信号转导; 另一方面, TNF- α 能通过下调过氧化物酶体增殖物活化受体, 间接降低胰岛素敏感性, 促进 IR。

2.2 KC 引起 NASH 的发病机制

根据 NAFLD 发病机制的“二次打击”学说, 第一次打击是脂肪贮积或 IR, 第二次打击则是氧化应激和异常细胞因子作用导致肝脏坏死性炎性反应

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和纤维化。从脂肪肝到脂肪性肝炎的进展过程是前炎性因子与抗炎性因子不平衡,触发氧化应激和脂质过氧化反应的结果。

2.2.1 KC与氧化应激 氧化应激在NAFLD中起重要作用,不断增强的氧应激及前炎性因子(如TNF- α)是导致NAFLD发病的“第二次打击”,导致肝脏坏死性炎症反应和纤维化。Koruk等^[8]发现,NASH患者血清中的氧化应激产物丙二醛(MDA)、NO、还原型谷胱甘肽和FFA明显增加,而抗氧化酶如超氧化物歧化酶(SOD)减少。在NASH大鼠模型中也发现类似结果,而且肝内KC活化与肝脏组织脂质过氧化损伤程度呈正相关。还有学者发现用GdCl₃预处理小鼠、大鼠,能明显减轻对乙酰氨基酚的肝毒性,NO和超氧化物生成减少^[9]。

此外,研究者在非酒精性脂肪肝患者的肝活组织切片中发现了一种壳质酶(CHIT)^[10],这种酶只在KC上表达,而且NAFLD患者的CHIT水平明显高于单纯性脂肪肝患者,CHIT的过度表达还可以激活肝星状细胞。

2.2.2 KC与内毒素及 β 细胞因子 目前已有几项临床研究显示NASH可被抗生素逆转,Wigg等^[11]进一步证实NASH患者存在着小肠内细菌过繁殖,可能在NASH的发病机制中发挥一定的作用。内毒素使KC激活,产生大量中间产物如前列腺素、活性氧、细胞因子(如TGF- β 、TNF- α 、IL-8)、NO及各种蛋白酶,从而诱导一系列类似于酒精性肝病表现。

2.2.3 KC与内源性乙醇 Cope等^[12]在NASH动物模型中发现胃肠源性乙醇,因此小肠内微生物可能通过产生乙醇发挥效应,其作用机制类似酒精性肝炎和慢性酒精性肝病。研究表明,急性或慢性乙醇注射都可以增加KC数量并激活KC,引起CD14表达上调,炎症介质IL-1、TNF- α 分泌增加,氧自由基产生增加。用GdCl₃特异性阻断KC后,可以逆转乙醇引起的肝脏炎症反应和肝细胞坏死。最近的研究还表明,慢性乙醇注射能降低KC细胞内cAMP水平并增强LPS对KC细胞NF- κ B通路的活化作用,引起TNF- α 生成增加^[13]。

2.2.4 KC与铁超负荷 1994年,Bacon首次发现许多NAFLD患者存在血清铁升高、铁超负荷(iron overload)现象,但具体机制不清。近年来在高脂饮食诱导的兔NASH模型上发现,肝内转化生长因子-1(TGF- β 1)、TNF- α 、IL-8和胶原 α 增加,红细胞脆性增加,细胞膜表面磷脂酰丝氨酸(PS)改变,使红细胞聚集,易在血流缓慢的肝窦内停滞,更易被

KC吞噬降解,红细胞血红蛋白中的铁导致肝脏内铁沉积^[14]。同时,铁超负荷又可以催化羟自由基产生、激活KC,激活NF- κ B和TNF- α 表达,诱导氧应激和脂质过氧化^[15]。

2.3 KC与NAFLD肝纤维化的发生机制

KC在NAFLD肝纤维化中的主要作用是在其激活后分泌细胞因子、产生氧化应激和/或促使星状细胞转化为成纤维细胞^[16-18]。肝星状细胞(HSC)是肝纤维化发展过程中的关键细胞。激活HSC的主要因素有氧化应激、转化生长因子-1(TGF- β 1)、结缔组织生长因子(CTGF)等。

研究比较NASH患者与单纯性脂肪肝患者的血清TGF- β 1水平后发现,TGF- β 1与纤维化发病机制关系密切。激活的KC可以分泌大量TGF- β 1,它是HSC和纤维母细胞的生长因子,而且是促使HSC活化的最强有力因子。CTGF在TGF- β 1的诱导下由HSC、成纤维细胞等间质细胞产生,有介导TGF- β 1对HSC促进其增生及细胞外基质合成的作用。研究观察到CTGF在NASH患者的肝组织内过表达,并与纤维化严重程度相关。另一项研究发现TNF- α 能诱导HSC中CTGF的表达。最近还有试验指出:与正常小鼠相比,TNF受体1(TNFR1)和2(TNFR2)基因双敲除小鼠给予不含蛋氨酸和胆酸的饮食后,肝内活化KC减少,TNF- α 含量降低,金属基质蛋白1(TIMPI)、细胞间黏附分子-1(ICAM-1)、血管细胞黏附分子-1(VCAM-1)都减少^[18-19]。

3 结语

随着生活水平的提高,肥胖群体的迅速扩增,我国NAFLD尤其是儿童及青少年NAFLD、NASH的发病率呈快速递增趋势,并有部分发生肝硬化^[20]。当前NAFLD缺乏特定有效的治疗方案,因此通过对NAFLD发病机制进一步的研究有助于防治NAFLD。既往许多研究已表明IR、细胞因子、氧化应激与NAFLD关系密切,然而其详细的作用机制以及KC在其中的具体作用机制都需要进一步研究。上述问题的解决,无疑将对NAFLD的防治发挥重要的推动作用。

参考文献

- 1 Marquez-Velasco R, Masso F, Hernandez-Pando R, et al. LPS pretreatment by the oral route protects against sepsis induced by cecal ligation and puncture. Regulation of proinflammatory response and IgM anti-LPS antibody production as associated mechanisms. *Inflamm Res*, 2007, 56: 385-390.
- 2 Dandona P, Aljada A, Chaudhuri A, et al. Metabolic syndrome: a comprehensive perspective based on interactions be-

- tween obesity, diabetes, and inflammation. *Circulation*, 2005, 111: 1448-1454.
- 3 Kelley DE, McKolanis TM. Fatty liver in type 2 diabetes mellitus: relation to regional adiposity, fatty acids, and insulin resistance. *Am J Physiol Endocrinol Metab*, 2003, 285: E906-E916.
 - 4 Marchesini G, Brizi M, Bianchi G, et al. Non-alcoholic fatty liver disease: a feature of the metabolic syndrome. *Diabetes*, 2001, 50: 1844-1850.
 - 5 De Taeye BM, Novitskaya T, McGuinness OP, et al. Macrophage TNF- α contributes to insulin resistance and hepatic steatosis in diet-induced obesity. *Am J Physiol Endocrinol Metab*, 2007, 293: E713-E725.
 - 6 Hotamisligil GS, Peraldi P, Budavari A, et al. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α and obesity-induced insulin resistance. *Science*, 1996, 271: 665-668.
 - 7 Diehl AM. Non-alcoholic steatosis and steatohepatitis IV. Non-alcoholic fatty liver disease abnormalities in macrophage function and cytokines. *Am J Physiol Gastrointest Liver Physiol*, 2002, 282: G1-G5.
 - 8 Koruk M, Taysi S, Savas MC, et al. Oxidative stress and enzymatic antioxidant status in patients with non-alcoholic steatohepatitis. *Ann Clin Lab Sci*, 2004, 34: 57-62.
 - 9 Younis HS, Parrish AR, Glenn Sipes I. The role of hepatocellular oxidative stress in Kupffer cell activation during 1,2-Dichlorobenzene-induced hepatotoxicity. *Toxicol Sci*, 2003, 76: 201-211.
 - 10 Malaguarnera L, Di Rosa M, Zambito AM, et al. Chitotriosidase gene expression in Kupffer cells from patients with non-alcoholic fatty liver disease. *Gut*, 2006, 55: 1313-1320.
 - 11 Wigg AJ, Roberts-Thomson IC, Dymock RB, et al. The role of small intestinal bacterial overgrowth, intestinal permeability, endotoxaemia, and tumour necrosis factor alpha in the pathogenesis of non-alcoholic steatohepatitis. *Gut*, 2001, 48: 206-211.
 - 12 Cope K, Risby T, Diehl AM. Increased gastrointestinal ethanol production in obese mice: implications for fatty liver disease pathogenesis. *Gastroenterology*, 2000, 119: 1340-1347.
 - 13 Mandrekar P. Signaling mechanisms in alcoholic liver injury: Role of transcription factors, kinases and heat shock proteins. *World J Gastroenterol*, 2007, 13: 4979-4985.

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- 6 Comlekci A, Akpınar H, Yesil S, et al. Serum leptin levels in patients with liver cirrhosis and chronic viral hepatitis. *Scand J Gastroenterol*, 2003, 38: 779-786.
- 7 Kalaitzakis E, Bosaeus I, Ohman L, et al. Altered postprandial glucose, insulin, leptin, and ghrelin in liver cirrhosis: correlations with energy intake and resting energy expenditure. *Am J Clin Nutr*, 2007, 85: 808-815.
- 8 Esler M, Vaz M, Collier G, et al. Leptin in human plasma is derived in part from the brain, and cleared by the kidney. *Lancet*, 1998, 351: 879.
- 9 Henriksen JH, Holst JJ, Møller S, et al. Increased circulating leptin in alcoholic cirrhosis: relation to release and disposal. *Hepatology*, 1999, 29: 1818-1824.
- 10 Honda H, Ikejima K, Hirose M, et al. Leptin is required for fibrogenic responses induced by thioacetamide in the murine liver. *Hepatology*, 2002, 36: 12-21.
- 11 Ikejima K, Takei Y, Honda H, et al. Leptin receptor-mediated signaling regulates hepatic fibrogenesis and remodeling of extracellular matrix in the rat. *Gastroenterology*, 2002, 122: 1399-1410.
- 12 Ikejima K, Okumura K, Kon K, et al. Role of adipocytokines in hepatic fibrogenesis. *J Gastroenterol Hepatol*, 2007, 22: 87-92.
- 13 Parsons CJ, Takashima M, Rippe RA. Molecular mechanisms of hepatic fibrogenesis. *J Gastroenterol Hepatol*, 2007, 22: 79-84.
- 14 Daniluk J, Szuster-Ciesielska A, Drabko J, et al. Serum cytokine levels in alcohol-related liver cirrhosis. *Alcohol*, 2001, 23: 29-34.
- 15 Odeh M, Sabo E, Srugo I, et al. Serum levels of tumor necrosis factor-alpha correlate with severity of hepatic encephalopathy due to chronic liver failure. *Liver Int*, 2004, 24: 110-116.
- 16 Lin SY, Wang YY, Sheu WH. Increased serum leptin concentrations correlate with soluble tumour necrosis factor receptor levels in patients with cirrhosis. *Clin Endocrinol*, 2002, 57: 805-811.
- 17 Mizuno TM, Funabashi T, Kleopoulos SP, et al. Specific preservation of biosynthetic responses to insulin in adipose tissue may contribute to hyperleptinemia in insulin-resistant obese mice. *Nutr*, 2004, 134: 1045-1450.
- 18 张予蜀, 袁捷, 张振玉, 等. 血清瘦素质量浓度与胃癌患者营养状态之间的关系. *东南大学学报医学版*, 2007, 26: 49-51.
- 19 张鸿, 王秋月, 侯刚, 等. 慢性阻塞性肺疾病患者血清瘦素水平与营养不良的关系. *中国医科大学学报*, 2006, 35: 168-170.
- 20 Yilmaz A, Kayardi M, Icagasioglu S, et al. Relationship between serum leptin levels and body composition and markers of malnutrition in nondiabetic patients on peritoneal dialysis or hemodialysis. *J Chin Med Assoc*, 2005, 68: 566-570.
- 21 Williams G, Harrold JA, Cutler DJ. The hypothalamus and the regulation of energy homeostasis: lifting the lid on a black box. *Proc Nutr Soc*, 2000, 59: 385-396.
- 22 何池义, 刘少锋, 张国政, 等. 肝硬化患者血清瘦素水平和营养状况的关系. *皖南医学院学报*, 2004, 23: 256-257.
- 23 焦秀娟, 姜慧卿, 彭勤, 等. 肝炎后肝硬化患者血清瘦素测定的临床意义. *中国实用内科杂志*, 2005, 25: 230-232.
- 24 Onodera K, Kato A, Suzuki K. Serum leptin concentrations in liver cirrhosis: relationship to the severity of liver dysfunction and their characteristic diurnal profiles. *Hepatol Res*, 2001, 21: 205-212.

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- cell in the human gastric corpus; a distinctive, functional cell lineage. *J Pathol*, 1999, 187;331-337.
- 2 Karam SM, Leblond CP. Dynamics of epithelial cells in the corpus of the mouse stomach. III. Inward migration of neck cells followed by progressive transformation into zymogenic cells. *Anat Rec*, 1993, 236;297-313.
 - 3 Karam SM. Dynamics of epithelial cells in the corpus of the mouse stomach. IV. Bidirectional migration of parietal cells ending in their gradual degeneration and loss. *Anat Rec*, 1993, 236;314-332.
 - 4 Dixon MF. Prospects for intervention in gastric carcinogenesis; reversibility of gastric atrophy and intestinal metaplasia. *Gut*, 2001, 49;2-4.
 - 5 Brembeck FH, Schreiber FS, Deramaudt TB, et al. The mutant K-ras oncogene causes pancreatic periductal lymphocytic infiltration and gastric mucous neck cell hyperplasia in transgenic mice. *Cancer Res*, 2003, 63;2005-2009.
 - 6 Yoshikazu K, Shunji I. Mechanism of gastric mucosal proliferation induced by gastrin. *J Gastroenterol Hepatol*, 2000, 15(Suppl): 7-11.
 - 7 Yonemura Y, Takashima T, Miwa K, et al. Amelioration of diabetes mellitus in partially depancreatized rats by poly(ADP-ribose) synthetase inhibitors. *Diabetes*, 1984, 263;2111-2114.
 - 8 Hartupce JC, Zhang H, Bonaldo MF, et al. Isolation and characterization of a cDNA encoding a novel member of the human regenerating protein family; RegIV. *Biochim Biophys Acta*, 2001, 1518;287-293.
 - 9 Watanabe T, Yonekura H, Terazono K, et al. Complete nucleotide sequence of human reg gene and its expression in normal and tumoral tissues. *J Biol Chem*, 1990, 265;7432-7439.
 - 10 Miyaoka Y, Kadowaki Y, Ishihara S, et al. Transgenic overexpression of Reg protein caused gastric cell proliferation and differentiation along parietal cell and chief cell lineages. *Oncogene*, 2004, 23;3572.
 - 11 Kazumori H, Ishihara S, Fukuda R, et al. Localization of Reg receptor in rat fundic mucosa. *J Lab Clin Med*, 2002, 139;101-108.
 - 12 Chiharu K, Hirokazu F, Yoshikazu K, et al. Regenerating gene expression in normal gastric mucosa and indomethacin-induced mucosal lesions of the rat. *J Gastroenterol*, 1997, 32;12-18.
 - 13 Fukui H, Fujii S, Takeda J, et al. Expression of regI alpha protein in human gastric cancers. *Digestion*, 2004;69;177-184.
 - 14 Tsutomu C, Hirokazu F, Yoshikazu K, et al. Reg protein: a possible mediator of gastrin-induced mucosal cell growth. *J Gastroenterol*, 2000, 35;52-56.
 - 15 Sekikawa A, Fukui H, Fujii S, et al. REGI alpha protein may function as a trophic and/or antiapoptotic factor in the development of gastric cancer. *Gastroenterology*, 2005, 128;642-653.
 - 16 Fukui H, Franceschi F, Penland RL, et al. Effects of helicobacter pylori infection on the link between regenerating gene expression and serum gastrin levels in Mongolian gerbils. *Lab Invest*, 2003, 83;1777-1786.
 - 17 Teo VF, Louise MJ, Nhung VN, et al. Growth factors associated with gastric mucosal hypertrophy in autoimmune gastritis. *Am J Physiol Gastrointest Liver Physiol*, 2004, 287;G910-918.
 - 18 Bockman DE, Sharp R, Merlino G, et al. Regulation of terminal differentiation of zymogenic cells by transforming growth factor α in transgenic mice. *Gastroenterology*, 1995, 108;447 - 454.
 - 19 Playford RJ, Boulton R, Ghatei MA, et al. Comparison of the effects of transforming growth factor alpha and epidermal growth factor on gastrointestinal proliferation and hormone release. *Digestion*, 1996, 57;362-367.
 - 20 袁庆丰, 姒健敏, 周雯, 等. 表皮生长因子促细胞增殖的量-效关系及对萎缩性胃炎大鼠胃粘膜的保护作用. *中华老年医学杂志*, 2006, 25;459-462.
 - 21 Masakyo A, Sotaro M, Shoichi S, et al. Reg gene expression is increased in rat gastric enterochromaffin-like cells following water immersion stress. *Gastroenterology*, 1996, 111;45-55.
 - 22 Hideaki K, Shunji I, Eiichi H, et al. Neutrophil chemoattractant 2β regulations expression of the Reg gene in injured gastric mucosa in rats. *Gastroenterology*, 2000, 119;1610-1622.

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(本文编辑:王立明)

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- 14 Otagawa K, Kinoshita K, Fujii H, et al. Erythrophagocytosis by liver macrophages (Kupffer cells) promotes oxidative stress, inflammation, and fibrosis in a rabbit model of steatohepatitis; implications for the pathogenesis of human non-alcoholic steatohepatitis. *Am J Pathol*, 2007, 170;967-980.
- 15 Tsukamoto H. Iron regulation of hepatic macrophage TNF- α expression. *Free Radic Biol Med*, 2002, 32;309-313.
- 16 Kawada N, Otagawa K. Role of oxidative stress and Kupffer cells in hepatic fibrosis. *J Gastroenterol Hepatol*, 2007, 22; S85-S86.
- 17 Esposito K, Giugliano D. Diet and inflammation; a link to metabolic and cardiovascular diseases. *Eur Heart J*, 2006, 27;15-20.
- 18 Lambeth JD. Nox enzymes, ROS, and chronic disease; an example of antagonistic pleiotropy. *Free Radic Biol Med*, 2007, 43; 332-347.
- 19 Tomita K, Tamiya G, Ando S, et al. Tumour necrosis factor α signalling through activation of Kupffer cells plays an essential role in liver fibrosis of non-alcoholic steatohepatitis in mice. *Gut*, 2006, 55;415-424.
- 20 Burke A, Lucey MR. Non-alcoholic fatty liver disease, non-alcoholic steatohepatitis and orthotopic liver transplantation. *Am J Transplant*, 2004, 4;686-693.

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