

## • 综述 •

# 幽门螺杆菌耐药的分子机制研究现状

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**摘要:**近年抗生素耐药率普遍升高,耐药成为根除幽门螺杆菌治疗失败的主要原因。克拉霉素耐药的主要原因为23S rRNA基因点突变。甲硝唑低水平耐药主要与硝基还原酶基因 $rdxA$ 和 $frxA$ 基因的失活有关,高水平耐药是多基因突变的累积效果所致。阿莫西林耐药与 $pbp1A$ 基因多位点突变相关。 $porD$ 和 $oorD$ 两基因的突变可能与幽门螺杆菌对呋喃唑酮产生低水平耐药相关。

**关键词:**幽门螺杆菌;耐药性;突变

目前已明确幽门螺杆菌(*Hp*)是慢性活动性胃炎的直接病因,与消化性溃疡、黏膜相关性淋巴样组织淋巴瘤密切相关,是胃癌发生的危险因子。根除*Hp*治疗是临床治疗慢性胃病的重要策略,根除*Hp*不仅能加速消化性溃疡的愈合,而且能显著降低溃疡复发率,也可以减少早期胃癌术后的复发。目前,阿莫西林、甲硝唑和克拉霉素是临幊上根除*Hp*治疗最常选用的抗生素,但随着抗生素的广泛应用,*Hp*的耐药率逐年上升<sup>[1]</sup>。国内不同地区、不同民族人群胃内*Hp*检出率在30%~80%之间,有很大差别。因此,*Hp*耐药机制的研究在临幊诊断和治疗中意义重大。

## 1 克拉霉素耐药的分子机制

克拉霉素是新一代的大环内酯类抗生素,其通过抑制细菌蛋白质的合成,达到杀菌目的。近年来*Hp*对克拉霉素的耐药率呈上升趋势,有文献报道,日本地区*Hp*对克拉霉素耐药率约13%~18%,欧洲约1.7%~23.4%,北美地区约12.2%~25%<sup>[2]</sup>。

目前对克拉霉素耐药机制的研究主要集中在对*Hp*23S rRNA基因的分析。目前为止已证实23SrRNA基因主要有3种不同的点突变与*Hp*对克拉霉素耐药有关:A2143G、A2142G、A2142C,其中最常见为A2143G,其次为A2142G,而A2142C则较少见。已有多项研究证实,A2142G和A2143G两种突变可以致*Hp*对克拉霉素高水平耐药,其中A2142G/C的平均最低抑菌浓度(MIC)较A2143G高,A2142G和A2143G的联合突变较单一突变MIC更高<sup>[2]</sup>。但并非所有A2142G突变MIC均较A2143G高,在Versalovic等的研究中,26.3%的

A2142G并非高水平耐药,也有研究发现2株A2142G的MIC较A2143G低,说明A2142G突变并非高水平耐药所必需<sup>[3]</sup>。

其他已发现的较少见突变包括A2115G-G2141A联合突变、T2717C、A2142T、T2182C、T2244C、G2223A、T2288C、C2195T-A2143G联合突变、T2221C-A2143G联合突变、G1939A、T1942C和C2147G三个位点联合突变<sup>[4]</sup>。Hulten于一株耐药株中发现A2115G-G2141A联合突变,而无A2143G、A2142G突变,证明A2115G-G2141A联合突变与*Hp*耐药相关<sup>[5]</sup>,其他位点与耐药的关系还有待于进一步研究。Rimbara等<sup>[4]</sup>将日本1995到2004年间检测的*Hp*临幊分离株进行META分析,发现A2142G或者A2143G突变出现于所有*Hp*耐克拉霉素菌株,敏感株则未检测出,T2182C、G2223A、T2288C、C2244T突变在敏感株和耐药株中均有较高的检出率,这4种突变与*Hp*耐克拉霉素无关。Matsuoka等<sup>[6]</sup>也证实T2182C、T2244C突变敏感株和耐药株均存在,与耐药无关。

由此可见,A2142G、A2143G、A2142C、A2115G-G2141A基因突变与*Hp*克拉霉素耐药相关,因前三者较常见,分子生物学方法检测克拉霉素耐药多立足于这三个基因突变位点,值得一提的是,有些学者检测克拉霉素耐药选择位点2182<sup>[7-8]</sup>,前面已提到Rimbara和Matsuoka已证实2182位点突变在敏感株和耐药株中均有较高的检出率,与耐药无关,故选择2182位点作为耐药检测位点显然是不合适的。

## 2 甲硝唑耐药的分子机制

甲硝唑耐药率是几种抗*Hp*药物中最高的,一些发展中国家达90%,西欧国家也达5%~50%<sup>[9]</sup>。甲硝唑耐药可能与 $rdxA$ 、 $frxA$ 、 $fdxB$ 基因突变有关。

$rdxA$ 和 $frxA$ 基因在甲硝唑耐药中所起的作用

基金项目:科技型中小企业技术创新基金资助项目(06C26214401607)  
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存在着争议。大部分学者认为,rdxA 基因失活突变是 *Hp* 甲硝唑耐药的基因基础。Moore 等<sup>[9]</sup>认为,frxA 基因的框移突变在敏感株和耐药株中出现的频率相同,因此与 *Hp* 甲硝唑耐药无关,只有 rdxA 基因与耐药相关。Jeong 等<sup>[10]</sup>认为,rdxA 的功能对于 *Hp* 对甲硝唑的敏感性起主要作用,临床分离株对甲硝唑的耐药基本上都有 rdxA 的突变,在 rdxA 基因突变的基础上,frxA 基因的失活突变可以增加 *Hp* 细菌的 MIC,frxA 基因的失活可以使甲硝唑的杀菌作用减缓,增加已有 rdxA 突变的 *Hp* 耐药水平,但在 rdxA 基因有活性的情况下不能使 *Hp* 产生耐药。rdxA 基因和 frxA 联合突变的耐药水平比 rdxA 单独突变的耐药性要高。Yang 等<sup>[11]</sup>证实 rdxA 基因突变对 *Hp* 甲硝唑耐药起主要作用,耐药株中 89% 存在 rdxA 基因突变,而敏感株中 rdxA 基因无突变而存在 frxA 基因突变,高水平耐药株中 86% 存在 rdxA 基因突变,只有 43% 存在 frxA 基因突变,表明 rdxA 基因突变在甲硝唑高水平耐药中所起的作用较 frxA 基因突变重要。

虽然 rdxA 基因的突变失活与甲硝唑耐药密切相关,很多研究显示,rdxA 基因治疗前后敏感株和耐药株的序列分析并无显著差别,说明 rdxA 基因非耐药必需基因,可能存在其他的机制。Marais 等<sup>[12]</sup>证明甲硝唑耐药可以只由 frxA 突变引起,也可以没有 rdxA 或 frxA 突变,存在其他耐药机制。目前的观点为,甲硝唑低水平的耐药(MTZ 32 μg/ml)是由于 rdxA 和 frxA 基因失活所致,高水平耐药很可能是多基因突变的累积效果。Albert 等<sup>[13]</sup>发明了一种以芯片杂交为基础的比较基因组测序法(CGS),用 CGS 来研究基因突变与 *Hp* 对甲硝唑耐药的关系,发现从敏感到高水平的耐药是从 rdxA 基因突变开始,接着是 frxA 基因突变,高水平耐药是由于多重基因突变叠加所致。

### 3 阿莫西林耐药的分子机制

阿莫西林又称羟氨苄西林是 *Hp* 根除治疗中常用的 β-内酰胺类抗生素,很多地区耐药率低于 1%<sup>[14]</sup>。PBP1 对阿莫西林亲和力下降可能是耐药的主要原因。近来研究涉及 PBP1 氨基酸序列和 pbp1 基因突变的具体位点之间关系。Paul 等<sup>[15]</sup>发现以下突变与阿莫西林耐药有关:pbp1(S414R、Y484C、T541I 和 P600T)。Gerrits 等<sup>[16]</sup>证明 Ser414Arg 在阿莫西林 Hardenberg strain 耐药株中起主要作用后进一步证实, S402G、E406A、S417T、S414R、T555S 和 N561Y 位点突变为耐药

主要原因。Rimbara 等<sup>[14]</sup>发现 9 个突变位点仅发生于耐药株和低度敏感株,其中 Asn562→Tyr 突变出现于所有阿莫西林耐药株中,为阿莫西林耐药株共同而特异的突变。Kwon 等<sup>[17]</sup>通过序列比对分析发现耐药株 pbp1-A 基因发生 51 个碱基突变,将其转入敏感株使其 MIC 升至 8 g/ml,而其他报道 pbp1-A 有 1 到 4 个位点突变时 MIC 为 2 g/ml。

高水平耐药是多基因多位点共同作用的结果,pbp1 基因突变并不能造成阿莫西林高水平耐药,阿莫西林高水平耐药存在 pbp1 突变以外的其他机制。Co 等<sup>[18]</sup>实验证明, pbp1-T438M、HopB、Hop 基因突变均参与阿莫西林耐药。Rimbara 等<sup>[19]</sup>实验证明 pbp1、pbp2, 和 pbp3 突变均与阿莫西林耐药相关,在 pbp1 突变的基础上 pbp2 和 pbp3 的突变更进一步提高了 MIC。

### 4 呋喃唑酮耐药的分子机制

呋喃唑酮近年来被越来越多地用于 *Hp* 感染的一线治疗,同样导致耐药出现。呋喃唑酮与甲硝唑作用机制类似,Kwon<sup>[20]</sup>推测 *Hp* 对呋喃唑酮的低水平耐药可能与 por 和 oor 两基因突变有关。Masaoka 等<sup>[21]</sup>发现呋喃唑酮耐药株 porD 基因有三个位点突变(G353A, A356G, 和 C357T),以 C357T 为常见,oorD 突变位点为 A041G, A122G, C349A (G),首先在国内外用数据支持了 Kwon 等的推测。

### 5 展望

*Hp* 是胃肠疾病的重要病原体,清除 *Hp* 对胃肠疾病的转归有重要作用。耐药基因突变检测若能成为 *Hp* 检查的常规检测,将会为临床与 *Hp* 相关性疾病的诊治提供依据。虽然目前国内对外对 *Hp* 耐药性的研究取得了一定进展,但仍存在着耐药机制的不同观点,基于 *Hp* 耐药基因突变检测的分子生物学检测方法存在分子基础上的不足,因此今后的研究方向重点将是分子机制与耐药性之间的关系。

### 参 考 文 献

- 1 Malfertheiner P, Megraud F, O'Morain C, et al. Current concepts in the management of Helicobacter pylori infection: the Maastricht III Consensus Report. Gut, 2007, 56: 772-781.
- 2 Nakamura A, Furuta T, Shirai N, et al. Determination of mutations of the 23S rRNA gene of Helicobacter pylori by allele specific primer-polymerase chain reaction method. J Gastroenterol Hepatol, 2007, 22: 1057-1063.
- 3 Furuta T, Soya Y, Sugimoto M, et al. Modified allele-specific primer-polymerase chain reaction method for analysis of susceptibility of Helicobacter pylori strains to clarithromycin. J Gastroenterol Hepatol, 2007, 22: 1810-1815.

- 4 Rimbara E, Noguchi N, Kijima H, et al. Mutations in the 23S rRNA gene of clarithromycin-resistant *Helicobacter pylori* from Japan. *Int J Antimicrob Agents*, 2007, 30: 250-254.
- 5 Hulten K, Gibreel A, Skold O, et al. Macrolide resistance in *Helicobacter pylori*: mechanism and stability in strains from clarithromycin-treated patients. *Antimicrob Agents Chemother*, 1997, 41: 2550-2553.
- 6 Matsuoka M, Yoshida Y, Hayakawa K, et al. Simultaneous colonisation of *Helicobacter pylori* with and without mutations in the 23S rRNA gene in patients with no history of clarithromycin exposure. *Gut*, 1999, 45: 503-507.
- 7 Chen S, Li Y, Yu C. Oligonucleotide microarray: a new rapid method for screening the 23S rRNA gene of *Helicobacter pylori* for single nucleotide polymorphisms associated with clarithromycin resistance. *J Gastroenterol Hepatol*, 2008, 23: 126-131.
- 8 Posteraro P, Branca G, Sanguinetti M, et al. Rapid detection of clarithromycin resistance in *Helicobacter pylori* using a PCR-based denaturing HPLC assay. *J Antimicrob Chemother*, 2006, 57: 71-78.
- 9 Moore JM, Salama NR. Mutational analysis of metronidazole-resistance in *Helicobacter pylori*. *Antimicrob Agents Chemother*, 2005, 49: 1236-1237.
- 10 Jeong JY, Mukhopadhyay AK, Akada JK, et al. Roles of FrxA and RdxA nitroreductases of *Helicobacter pylori* in susceptibility and resistance to metronidazole. *J Bacteriol*, 2001, 183: 5155-5162.
- 11 Yang YJ, Wu JJ, Sheu BS, et al. The rdxA gene plays a more major role than frxA gene mutation in high-level metronidazole resistance of *Helicobacter pylori* in Taiwan. *Helicobacter*, 2004, 9: 400-407.
- 12 Marais A, Bilardi C, Cantet F, et al. Characterization of the genes rdxA and frxA involved in metronidazole resistance in *Helicobacter pylori*. *Res Microbiol*, 2003, 154: 137-144.
- 13 Albert TJ, Dailidiene D, Dailide G, et al. Mutation discovery in bacterial genomes: metronidazole resistance in *Helicobacter pylori*. *Nat Methods*, 2005, 2: 951-953.
- 14 Rimbara E, Noguchi N, Kawai T, et al. Correlation between substitutions in penicillin-binding protein 1 and amoxicillin resistance in *Helicobacter pylori*. *Microbiol Immunol*, 2007, 51: 939-944.

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- 3 Mesri M, Morales-Ruiz M, Ackermann EJ, et al. Suppression of vascular endothelial growth factor-mediated endothelial cell protection by Survivin targeting. *Am J Pathol*, 2001, 158: 1757-1765.
- 4 Sui L, Dong Y, Watanabe Y, et al. Alteration and clinical relevance of PTEN expression and its correlation with Survivin expression in epithelial ovarian tumors. *Oncol Rep*, 2006, 15: 773-778.
- 5 Bao ST, Gui SQ, Lin MS. Relationship between expression of Smac and Survivin and apoptosis of primary hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int*, 2006, 5: 580-583.
- 6 Morinaga S, Nakamura Y, Ishiwa N, et al. Expression of Survivin mRNA associates with apoptosis, proliferation and histologically aggressive features in hepatocellular carcinoma. *Oncol Rep*, 2004, 12: 1189-1194.
- 7 Ye CP, Qiu CZ, Huang ZX, et al. Relationship between Survivin expression and recurrence, and prognosis in hepatocellular carcinoma. *World J Gastroenterol*, 2007, 13: 6264-6268.
- 8 Zamecnik PC, Stephenson ML. Inhibition of Rous sarcoma virus replication and cell transformation by a specific oligodeoxy-nucleotide. *Proc Natl Acad Sci U S A*, 1978, 75: 280-284.
- 9 Glodde M, Sirsi SR, Lutz GJ. Physicochemical properties of low and high molecular weight poly(ethylene glycol)-grafted poly(ethylene imine) copolymers and their complexes with oligonucleotides. *Biomacromolecules*, 2006, 7: 347-356.
- 10 Altieri DC. Validating Survivin as a cancer therapeutic target. *Nat Rev Cancer*, 2003, 3: 46.
- 11 Chen J, Wu W, Tahir SK, et al. Down-regulation of Survivin by antisense oligonucleotides increases apoptosis, inhibits cyto-kinetics and anchorage-independent growth. *Neoplasia*, 2000, 2: 235-241.
- 12 Chen T, Jia YR, Zhao TJ, et al. Inhibitory effect of antisense oligonucleotide on the expression of Survivin gene and proliferation of human hepatocellular carcinoma cell line SMMC-7721. *World Chin J Digestol*, 2004, 12: 1546-1549.
- 13 Dai DJ, Lu CD, Lai RY, et al. Survivin antisense compound inhibits proliferation and promotes apoptosis in liver cancer cells. *World J Gastroenterol*, 2005, 11: 193-199.
- 14 高鹏, 方驰华, 张刚庆. Survivin 反义寡核苷酸抑制肝癌裸鼠皮下移植瘤生长的作用. 中国病理生理杂志, 2008, 24: 101-104.
- 15 Lefranc F, Facchini V, Kiss R, et al. Proautophagic drugs: a novel means to combat apoptosis-resistant cancers, with a special emphasis on glioblastomas. *Oncologist*, 2007, 12: 1395-1403.
- 16 Chang Q, Liu ZR, Wang DY, et al. Survivin expression induced by doxorubicin in cholangiocarcinoma. *World J Gastroenterol*, 2004, 10: 415-418.
- 17 Tamm I, Dürken B, Hartmann G. Antisense therapy in oncology: new hope for an old idea? *Lancet*, 2001, 358: 489-497.
- 18 Gao P, Fang CH, Zhang GQ, et al. Antisense oligonucleotide against Survivin induces apoptosis and enhances adriamycin sensitivity of SMMC-7721/ADM cells. *Nan Fang Yi Ke Da Xue Xue Bao*, 2006, 26: 1644-1647.
- 19 王颖, 王家骁. 生存素反义寡核苷酸诱导肝癌细胞凋亡的实验研究. 中华消化杂志, 2003, 23: 11-14.
- 20 张鹤文, 汤恢焕, 王志明, 等. 靶向抑制 Survivin 基因对肝细胞癌血管生成的影响. 中华普通外科杂志, 2007, 22: 133-136.

(收稿日期: 2008-08-12)

(本文编辑: 李瑞芳)

- ERK5 signalling pathway. *Cell Signal*, 2006, 18: 753-760.
- 2 Weston CR, Davis RJ. The JNK signal transduction pathway. *Curr Opin Cell Biol*, 2007, 19: 142-149.
  - 3 Davis RJ. Signal transduction by the JNK group of MAP kinases. *Cell*, 2000, 103: 239-252.
  - 4 Swantek JL, Cobb MH, Geppert TD. Jun N-terminal kinase/stress-activated protein kinase (JNK/SAPK) is required for lipopolysaccharide stimulation of tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) translation: glucocorticoids inhibit TNF- $\alpha$  translation by blocking JNK/SAPK. *Mol Cell Biol*, 1997, 17: 6274-6282.
  - 5 Ma W, Gee K, Lim W, et al. Dexamethasone inhibits IL-12p40 production in lipopolysaccharide-stimulated human monocytic cells by down-regulating the activity of c-Jun N-terminal kinase, the activation protein-1, and NF- $\kappa$ B transcription factors. *J Immunol*, 2004, 172: 318-330.
  - 6 Saatian B, Zhao Y, He D, et al. Transcriptional regulation of lysophosphatidic acid-induced interleukin-8 expression and secretion by p38 MAPK and JNK in human bronchial epithelial cells. *Biochem J*, 2006, 393: 657-668.
  - 7 Pedra JH, Mattner J, Tao J, et al. c-Jun NH<sub>2</sub>-terminal kinase 2 inhibits gamma interferon production during anaplasma phagocytophilum infection. *Infect Immun*, 2008, 76: 308-316.
  - 8 Hoffmann E, Ashouri J, Wolter S, et al. Transcriptional regulation of EGR-1 by the interleukin-1-JNK-MKK7-c-Jun pathway. *J Biol Chem*, 2008, 283: 12120-12128.
  - 9 Yang H, Young DW, Gusovsky F, et al. Cellular events mediated by lipopolysaccharide-stimulated toll-like receptor 4. MD-2 is required for activation of mitogen-activated protein kinases and Elk-1. *J Biol Chem*, 2000, 275: 20861-20866.
  - 10 Adhikary G, Sun Y, Pearlman E. C-Jun NH<sub>2</sub> terminal kinase (JNK) is an essential mediator of Toll-like receptor 2-induced corneal inflammation. *J Leukoc Biol*, 2008, 83: 991-997.
  - 11 Abreu MT, Arnold ET, Thomas LS, et al. TLR4 and MD-2 expression is regulated by immune-mediated signals in human intestinal epithelial cells. *J Biol Chem*, 2002, 277: 20431-20437.
  - 12 Dong C, Yang DD, Wysk M, et al. Defective T cell differentiation in the absence of Jnk1. *Science*, 1998, 282: 2092-2095.
  - 13 Gao M, Labuda T, Xia Y, et al. Jun turnover is controlled through JNK dependent phosphorylation of the E3 ligase Itch. *Science*, 2004, 306: 271-275.
  - 14 Yang DD, Conze D, Whitmarsh AJ, et al. Differentiation of CD4 $^{+}$  T cells to Th1 cells requires MAP kinase JNK2. *Immunity*, 1998, 9: 575-585.
  - 15 Conze D, Krahl T, Kennedy N, et al. c-Jun NH(2)-terminal kinase (JNK)1 and JNK2 have distinct roles in CD8(+) $T$  cell activation. *J Exp Med*, 2002, 195: 811-823.
  - 16 Salh B. C-Jun N-terminal kinases as potential therapeutic targets. *Expert Opin Ther Targets*, 2007, 11: 1339-1353.
  - 17 Han Z, Boyle DL, Chang L, et al. c-Jun N-terminal kinase is required for metalloproteinase expression and joint destruction in inflammatory arthritis. *J Clin Invest*, 2001, 108: 73-81.
  - 18 Waetzig GH, Seegert D, Rosenstiel P, et al. p38 mitogen-activated protein kinase is activated and linked to TNF- $\alpha$  signaling in inflammatory bowel disease. *J Immunol*, 2002, 168: 5342-5351.
  - 19 Hommes D, van den Blink B, Plasse T, et al. Inhibition of stress-activated MAP kinases induces clinical improvement in moderate to severe Crohn's disease. *Gastroenterology*, 2002, 122: 7-14.
  - 20 Mitsuyama K, Suzuki A, Tomiyasu N, et al. Pro-inflammatory signaling by Jun-N-terminal kinase in inflammatory bowel disease. *Int J Mol Med*, 2006, 17: 449-455.
  - 21 Assi K, Pillai R. The specific JNK inhibitor SP600125 targets tumour necrosis factor- $\alpha$  production and epithelial cell apoptosis in acute murine colitis. *Immunology*, 2006, 118: 112-121.
  - 22 Malamut G, Cabane C, Dubuquoy L, et al. No evidence for an involvement of the p38 and JNK mitogen-activated protein in inflammatory bowel diseases. *Dig Dis Sci*, 2006, 51: 1443-1453.
  - 23 Chromik AM, Muller AM, Korner J, et al. Genetic deletion of JNK1 and JNK2 aggravates the DSS-induced colitis in mice. *J Invest Surg*, 2007, 20: 23-33.

(收稿日期: 2008-10-29)

(本文编辑:周骏)

## (上接第 121 页)

- 15 Paul R, Postius S, Melchers K, et al. Mutations of the Helicobacter pylori genes rdxA and pbp1 cause resistance against metronidazole and amoxicillin. *Antimicrob Agents Chemother*, 2001, 45: 962-965.
- 16 Gerrits MM, Godoy AP, Kuipers EJ, et al. Multiple mutations in or adjacent to the conserved penicillin-binding protein motifs of the penicillin-binding protein 1A confer amoxicillin resistance to Helicobacter pylori. *Helicobacter*, 2006, 11: 181-187.
- 17 Kwon DH, Dore MP, Kim JJ, et al. High-level beta-lactam resistance associated with acquired multidrug resistance in Helicobacter pylori. *Antimicrob Agents Chemother*, 2003, 47: 2169-2178.
- 18 Co EM, Schiller NL. Resistance mechanisms in an in vitro-selected amoxicillin-resistant strain of Helicobacter pylori. *Anti-microb Agents Chemother*, 2006, 50: 4174-4176.
- 19 Rimbara E, Noguchi N, Kawai T, et al. Mutations in penicillin-binding proteins 1, 2 and 3 are responsible for amoxicillin resistance in Helicobacter pylori. *J Antimicrob Chemother*, 2008, 61: 995-998.
- 20 Kwon DH, Lee M, Kim JJ, et al. Furazolidone- and nitrofurantoin-resistant Helicobacter pylori; prevalence and role of genes involved in metronidazole resistance. *Antimicrob Agents Chemother*, 2001, 45: 306-308.
- 21 Masaoka T, Suzuki H, Kurabayashi K, et al. Could frameshift mutations in the frxA and rdxA genes of Helicobacter pylori be a marker for metronidazole resistance? *Alimentary Pharmacology & Therapeutics*, 2006, 24: 81-87.

(收稿日期 2008-08-14)

(本文编辑:李瑞芳)